How Iodide Reaches its Site of Utilisation in the Thyroid Gland – Involvement of Solute Carrier 26A4 (Pendrin) and Solute Carrier 5A8 (Apical Iodide Transporter)

a report by
Bernard A Rousset

Professor of Cell Biology and Molecular Endocrinology, Lyon University Medical School DOI:10.17925/EE.2007.00.01.81

Iodine is an essential component of thyroid hormones thyroxine (T4) and triiodothyronine (T3), comprising 65 and 59% of their respective molecular weight. Thyroid hormones, and therefore iodine, are essential for mammalian life: they regulate many important biochemical reactions, especially the synthesis of proteins with key functional activities. Most dietary iodine is reduced to iodide before absorption through the gut and then transferred into the blood. The thyroid gland extracts and concentrates iodide from plasma. The normal thyroid maintains a concentration of free iodide 20–50 times higher than that of plasma, depending on the amount of available iodine and the activity of the gland. This concentration gradient may be more than 100:1 in a hyperactive thyroid, as seen in patients with Graves' disease. The daily intake of an adult human varies from less than 10mg in areas of extreme iodine deficiency to several hundred milligrams for some people receiving medicinal iodine. Too little iodine causes mental retardation, goitre, hypothyroidism and other features of the so-called iodine deficiency disorders (International Council for Control of Iodine Deficiency Disorders current iodine deficiency disorders status database – http://www.iccidd.org). Too much iodine increases the incidence of iodine-

> Thyroid hormones, and therefore iodine, are essential for mammalian life: they regulate many important biochemical reactions, especially the synthesis of proteins with key functional activities.

induced hyperthyroidism, autoimmune thyroid disease and perhaps thyroid cancer. The appropriate intake is about 150mg/day.

Iodide entering the thyroid gland must sequentially cross two lipid bilayers: the basolateral and then the apical plasma membrane of polarised thyroid epithelial cells or thyrocytes forming the functional units of the gland, the follicles. Thyroid follicles are made of a single layer of polarised cells delimiting a closed internal compartment. Once in the lumen of follicles, iodide is used to generate iodothyronine residues inside thyroglobulin molecules for the subsequent production of free thyroid hormones. As with other small charged molecules, iodide cannot easily diffuse through cell membranes; thus, its transfer from one side to the other of the thyroid cell plasma membrane requires a membrane protein.

The protein ensuring the active transport of iodide at the basolateral plasma membrane of thyrocytes (named NIS for Na+/iodide symporter) is now well characterised.[1] The movement of iodide from the cytoplasm of thyrocytes to the follicle lumen is expected to be a passive process; indeed,

due to the favourable electrochemical gradient, iodide efflux would require only a 'permease' or an ion channel. The transport of iodide from the cytoplasm of thyrocytes to the follicle lumen has been elegantly studied on polarised porcine thyrocytes cultured as tight monolayers in bicameral devices;[2] it was reported that the apical efflux of iodide, which is rapidly increased in response to thyroid-stimulating hormone (TSH), could occur through a cyclic adenosine monophosphate (cAMP) regulated 'iodide channel'. A study performed on thyroid membrane vesicles[3] also concluded that an iodide channel must exist in the thyroid. Molecular identification of the protein ensuring the transport of iodide at the apical membrane of thyrocytes has been the goal of several recent studies.

Pendrin, the protein encoded by the gene altered in Pendred's syndrome (PDS), i.e. the PDS gene, has been the first candidate. Pendrin, a membrane protein located at the apical pole of thyrocytes, is endowed with ion transport activity. Pendrin was rapidly considered as the apical iodide transporter despite the uncertainties and/or discrepancies that were apparent since the first reports. The initial proposal of pendrin as a sulphate transporter[4] was rapidly abandoned.[5] Most subsequent studies performed on different experimental systems assigned a function of anion exchanger to pendrin, exchanging chloride for hydroxide, bicarbonate or formate;[6–9] some studies based on transfection of recipient cells concluded that pendrin could be active as a chloride/iodide transporter[10,11] or chloride/iodide exchanger.[12]

It is difficult to envisage any physiological contribution from a chloride/iodide exchanger in the apical iodide efflux, since both luminal and cytoplasmic chloride concentrations are more than 1,000-fold greater than that of iodide; an exchange of cytoplasmic iodide with luminal chloride on a one-to-one stoechiometry seems unlikely. Still more confusing are the reports claiming that pendrin mediates iodide uptake (not efflux) by MCF-7 cells.[13] At present, there are no experimental data showing that pendrin activity accounts for the 'transfer' of iodide from thyrocytes to the apical compartment in a polarised thyroid cell system. Data from polarised Madin-Darby canine kidney cells transfected with the PDS gene,[14] although interesting, should not be extrapolated and considered as a demonstration of the function of pendrin in the thyroid. Several other arguments undermine the hypothesis of pendrin as an apical iodide transporter. First, the

Bernard A Rousset is Professor of Cell Biology and Molecular Endocrinology at the Lyon University Medical School. He is head of a research laboratory supported by the French Institute of Health and Medical Research, the Inserm Unit 369. His own research activities, previously centred on cell–cell communication processes in the thyroid gland, are now mainly directed towards studies of the functional genomics of thyroid cancer. Since the beginning of his career, he has been interested in iodine metabolism and hormonogenesis of the thyroid gland.

knockout of the PDS gene in the mouse, which causes functional alterations in the ear, does not induce any thyroid phenotype.[15]

> Indecision about the identity of the apical iodide transporter has been complicated by a report describing another candidate named apical iodide transporter.

Second, it has often been found that many patients with Pendred's syndrome who have a non-functional pendrin do not exhibit any thyroid alteration. Third, it seems difficult to accept that a given protein could have different functions in different organs. In the thyroid, pendrin – SLC26A4 in the solute carrier (SLC) transporter nomenclature – could exert a function of chloride/bicarbonate exchange (as in the kidney) and play, for example, a role in the control of luminal pH with a possible incidence on iodide oxidation.

Indecision about the identity of the apical iodide transporter has been complicated by a report describing another candidate named apical iodide transporter (AIT).[16] As with pendrin, AIT appears selectively located at the apical plasma membrane of thyrocytes and, as with pendrin, was reported to cause iodide discharge in transfected cell systems. The first problem arose when it was found, using colon cell lines, that this protein – structurally related to NIS – was an Na+-dependent transporter[17] belonging to the SLC5 family as the SLC5A8 member. Indeed, both previous studies and prediction on biological considerations do not point to the requirement for an Na+ dependency for the apical iodide transport. Soon after, two independent groups described SLC5A8 as an Na+-coupled transporter for short-chain fatty acids.[18–20] Recent studies on SLC5A8 indicate a consensus on this function.[21–23] Attempts to detect an activity of SLC5A8 in iodide efflux have been unsuccessful.[23]

At this stage, one has to conclude that the identity of the protein(s) ensuring the apical iodide transport in the thyroid is not known. ■

1. Dohan O, De la Vieja A, Paroder V, et al., *Endocr Rev*, 2003;24:48–77.
2. Nilsson M, Bjorkman U, Ekholm R, Ericson LE, *Eur J Cell Biol*, 1990;52:270–81.
3. Golstein P, Abramow M, Dumont JE, et al., *Am J Physiol*, 1992;263:C590–97.
4. Everett LA, Glaser B, Beck JC, et al., *Nat Genet*, 1997;17: 411–22.
5. Kraiem Z, Heinrich R, Sadeh O, et al., *J Clin Endocrinol Metab*, 1999;84:2574–6.
6. Scott DA, Karniski LP, *Am J Physiol Cell Physiol*, 2000;278:C207–11.
7. Soleimani M, *Curr Opin Nephrol Hypertens*, 2001;10:677–83.
8. Soleimani M, Greeley T, Petrovic S, et al., *Am J Physiol Renal Physiol*, 2001;280:F356–64.
9. Quentin F, Chambrey R, Trinh-Trang-Tan MM, et al., *Am J Physiol Renal Physiol*, 2004;287:F1179–88.
10. Scott DA, Wang R, Kreman, et al., *Nat Genet*, 1999;21:440–43.
11. Yoshida A, Taniguchi S, Hisatome I, et al., *J Clin Endocrinol Metab*, 2002;87:3356–61.
12. Yoshida A, Hisatome I, Taniguchi S, et al., *Endocrinology*, 2004;145:4301–8.
13. Rillema JA, Hill MA, *Exp Biol Med*, 2003;228:1078–82.
14. Gillam MP, Sidhaye AR, Lee EJ, et al., *J Biol Chem*, 2004;279:13004–10.
15. Everett LA, Belyantseva IA, Noben-Trauth K, et al., *Hum Mol Genet*, 2001;10:153–61.
16. Rodriguez AM, Perron B, Lacroix L, et al., *J Clin Endocrinol Metab*, 2002;87:3500–3.
17. Li H, Myeroff L, Smiraglia D, et al., *Proc Natl Acad Sci U S A*, 2003;100:8412–17.
18. Miyauchi S, Gopal E, Fei YJ, Ganapathy V, *J Biol Chem*, 2004;279:13293–6.
19. Coady MJ, Chang MH, Charron FM, et al., *J Physiol*, 2004;557: 719–31.
20. Gopal E, Fei YJ, Sugawara M, et al., *J Biol Chem*, 2004;279:44522–32.
21. Ganapathy V, Gopal E, Miyauchi S, et al., *Biochem Soc Trans*, 2005;33:237–40.
22. Gupta N, Martin PM, Prasad PD, et al., *Life Sci*, 2006;78:2419–25.
23. Paroder V, Spencer SR, Paroder M, et al., *Proc Natl Acad Sci U S A*, 2006;103:7270–75.

The following articles relating to thyroid disorders can be found on the website supporting the Touch Briefings healthcare series – www.touchbriefings.com