Anticancer Drug-induced Thyroid Dysfunction

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Cancer immunotherapy and targeted therapy, though less toxic than conventional chemotherapy, can increase the risk of thyroid dysfunction. Immune checkpoint inhibitors render the cancer cells susceptible to immune destruction, but also predispose to autoimmune disorders like primary hypothyroidism as well as central hypothyroidism secondary to hypophysitis. Tyrosine kinase inhibitors act by blocking vascular endothelial growth factor receptors and their downstream targets. Disruption of the vascular supply from the inhibition of endothelial proliferation damages not only cancer cells but also organs with high vascularity like the thyroid. Interferon-α, interleukin-2 and thalidomide analogues can cause thyroid dysfunction by immune modulation. Alemtuzumab, a monoclonal antibody directed against the cell surface glycoprotein CD52 causes Graves’ disease during immune reconstitution. Metaiodobenzylguanidine, combined with 131-Iodine, administered as a radiotherapeutic agent for tumours derived from neural crest cells, can cause primary hypothyroidism. Bexarotene can produce transient central hypothyroidism by altering the feedback effect of thyroid hormone on the pituitary gland. Thyroid dysfunction can be managed in the usual manner without a requirement for dose reduction or discontinuation of the implicated agent. This review aims to highlight the effect of various anticancer agents on thyroid function. Early recognition and appropriate management of thyroid disorders during cancer therapy will help to improve treatment outcomes.

Keywords
Thyroid, hypothyroidism, anticancer drugs, immune checkpoint inhibitors, tyrosine kinase inhibitors

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The armamentarium of anticancer drugs available to an oncologist has grown rapidly over the past few decades. The use of cancer immunotherapy and targeted therapy has become more popular in the last few years. It has also become increasingly clear that various anticancer agents, both conventional and newer ones, may be associated with certain off-target adverse effects involving the endocrine system, especially the thyroid gland.¹ The site of action of commonly used cancer immunotherapy agents is depicted in Figure 1. This article is aimed at describing thyroid dysfunction associated with various anticancer agents. These have been briefly summarised in Table 1. It is important to identify and appropriately treat thyroid dysfunction in such patients. This will not only improve their overall quality of life, but also ensure optimal treatment outcome.

Literature search
A MEDLINE search was conducted for articles published before 30 April 2019. Articles published in English were considered. The search terms were words related to thyroid disorders, such as ‘thyroid’, ‘hypothyroidism’, ‘thyrotoxicosis’, ‘hyperthyroidism’ ‘Graves’ disease’ ‘central hypothyroidism’, and ‘thyroiditis’ in association with ‘anticancer drugs’, ‘immune checkpoint inhibitors’, ‘tyrosine kinase inhibitors’, ‘interferon-α’, ‘interleukin-2’, ‘alemtuzumab’, ‘thalidomide’, ‘lenalidomide’, ‘pomalidomide’ ‘radioiodine-based cancer therapies’ and ‘bexarotene’. The names of specific drugs, like ipilimumab, nivolumab and pembrolizumab amongst immune checkpoint inhibitors; and sunitinib, sorafenib, axitinib, pazopanib, vandetanib, motesanib, imatinib, cabozantinib, nilotinib, dasatinib, erlotinib, gefitinib, lapatinib, nintedanib, regorafenib and tivozanib amongst tyrosine kinase inhibitors, were also included in the search. The abstracts were evaluated for relevance, and the full text of all appropriate articles was retrieved. Reference lists of selected articles were also searched. Articles describing usage of anticancer agents for the treatment of thyroid cancer were excluded.

Immune checkpoint inhibitors
A better understanding of the complexities of the human immune system and its regulation paved the way for a novel concept in the field of oncology termed ‘cancer immunotherapy’. The basic principle of cancer immunotherapy is utilisation of the body’s own immune system to target cancer cells. Immune checkpoint molecules are regulators of the immune system which provide self-tolerance and prevent the immune system from destroying its own cells (Figure 2). These include cytotoxic T-lymphocyte associated protein 4 (CTLA-4) and programmed cell death protein 1 (PD-1; a cell surface receptor) and its ligand (PD-L1). CTLA-4 is constitutionally expressed on regulatory T cells, gets up-regulated on T-cell activation, and works toward inhibiting a second (co-stimulatory) signal for T-cell activation. PD-1 is present on T cells, B cells and natural killer (NK) cells, and binds to PD-L1 expressed by tumour cells. The interaction between PD-1 and PD-L1 inhibits the destruction of tumour cells by the immune system; hence, PD-L1 is overexpressed by the tumour cells to their advantage.²³

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Hypothyroidism, ischaemic thyroiditis
Thalidomide analogues
Radioiodine-based cancer therapy
Bexarotene
Conventional agents

Indications and adverse events

Immune checkpoint inhibitors are agents that inhibit these immune checkpoint molecules, causing an immune attack on the tumour cells. The US Food and Drug Administration (FDA)-approved immune checkpoint inhibitors include anti-CTLA-4 monoclonal antibody (ipilimumab), anti-PD-1 monoclonal antibody (nivolumab, pembrolizumab), and anti-PD-L1 monoclonal antibody (atezolizumab, avelumab, and durvalumab). While initially approved for the treatment of unsectable or metastatic melanoma, these agents have been found to be effective in a wide variety of tumours, such as small-cell lung cancer, non-small-cell lung cancer (NSCLC), Hodgkin’s lymphoma, renal cell cancer (RCC), prostate cancer, bladder cancer, oesophageal cancer and breast cancer. The enhanced immune activation seen with these agents can lead to a variety of immune-related adverse events. These may involve a number of organ systems – skin (rash, pruritus), gastrointestinal tract and liver (colitis, autoimmune hepatitis), and the endocrine system (hypophysitis and thyroid dysfunction). In terms of endocrine dysfunction, hypophysitis has been reported more often with CTLA-4 inhibitors, while thyroid dysfunction has been reported with both CTLA-4 and PD-1/PD-L1 inhibitors.4

Primary thyroid dysfunction

Primary thyroid dysfunction following immune checkpoint-inhibitor therapy has been described in the form of subclinical or overt hypothyroidism, transient thyrotoxicosis and painless thyroiditis.6–8 Rare cases of Graves’ disease and euthyroid orbitopathy have also been described.6–10 The incidence of thyroid dysfunction following immune checkpoint-inhibitor therapy has been variably reported at 1–22% in the literature.6,7,10–11 The incidence is much higher in patients receiving a combination therapy (CTLA-4 inhibitor and PD-1 inhibitor). In the study by Ryder et al., the overall incidence of hypothyroidism/thyroiditis following ipilimumab was 6%, while it increased to 22% in those receiving a combination of ipilimumab and nivolumab.2

In the KEYNOTE-001 study, 51 subjects with advanced NSCLC treated with PD-1 inhibitor pembrolizumab were studied for thyroid dysfunction. Of the 48 subjects with euthyroidism at baseline, 10 (21%) developed hypothyroidism (requiring replacement) at a median duration of 6 weeks. Hypothyroidism was preceded by a phase of transient thyrotoxicosis in 6/10 (60%) subjects. Anti-thyroid antibodies were more frequent in

Hypophysitis and central hypothyroidism

Hypophysitis related to CTLA-4 inhibitor therapy has been reported at an incidence of 0.4–17.0% in literature.6–7 Similar to autoimmune lymphocytic hypophysitis, there is predilection for involvement of thyrotropes (central hypothyroidism) and corticotrophs (central hypo-adrenalism). However, unlike lymphocytic hypophysitis, CTLA-4 inhibitor-related hypophysitis is more common in males. A possible reason could be the higher proportion of male participants in studies reporting hypophysitis with CTLA-4-inhibitor therapy. In a retrospective study involving 154 adult subjects with melanoma treated with ipilimumab at a tertiary care centre in USA, hypophysitis was reported at a prevalence of 11% (17/154).4 The prevalence was significantly higher in males compared with females (15.6% versus 3.6%), and subjects with hypophysitis were significantly older compared to the remaining cohort (mean age 68.2 versus 59.9 years). The diagnosis of hypophysitis was made after a median duration of 8.4 weeks following the initiation of ipilimumab. Central hypothyroidism was reported in all, while central hypo-adrenalism was seen in 42% (7/17) subjects with hypophysitis. Interestingly, the median survival in subjects who developed hypophysitis was significantly higher than those who did not (19.4 versus 8.8 months). In another study by Ryder et al. (n=254), the overall incidence of hypophysitis following ipilimumab therapy was 8%, while in the subjects receiving combination immune checkpoint-inhibitor therapy (ipilimumab and nivolumab), the incidence increased to 9%.7

Table 1: Anticancer drugs causing thyroid dysfunction

<table>
<thead>
<tr>
<th>Immune checkpoint inhibitors</th>
<th>Hypophysitis (ipilimumab, nivolumab)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tyrosine kinase inhibitors</td>
<td>Hypothyroidism (sunitinib, sorafenib, axitinib, pazopanib, vandetanib, motesanib)</td>
</tr>
<tr>
<td>Interferon-α</td>
<td>Hypothyroidism, destructive thyroiditis</td>
</tr>
<tr>
<td>Interleukin-2</td>
<td>Hypothyroidism</td>
</tr>
<tr>
<td>Alemtuzumab</td>
<td>Graves’ disease</td>
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<tr>
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<tr>
<td>Bexarotene</td>
<td>Central hypothyroidism</td>
</tr>
<tr>
<td>Conventional agents</td>
<td>Alteration in thyroid binding proteins (not clinically relevant) (mitotane, 5-fluorouracil, oestrogens, tamoxifen, podophyllin, L-asparaginase)</td>
</tr>
</tbody>
</table>

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the subset of subjects who developed thyroid dysfunction (8/10, 80%), compared to those who did not (3/38, 8%). Similar to the ipilimumab hypophysitis study, subjects developing thyroid dysfunction had overall better survival (hazard ratio, 0.29; 95% confidence interval 0.09–0.94; \(p=0.04\)). The authors concluded that primary thyroid dysfunction following pembrolizumab therapy is fairly common, occurs early, is associated with transient thyrotoxicosis and anti-thyroid antibodies in most cases, and possibly portends a favourable outcome.

The association between immune checkpoint inhibitor-related endocrine dysfunction and better overall survival seen in various studies might indicate higher immune activation and better tumour-cell destruction by the immune system in subjects developing this adverse effect. This interesting observation needs further validation with well-designed, long-term follow-up studies.

**Management**

The mainstay of management of thyroid dysfunction following immune checkpoint inhibitor therapy involves appropriate thyroid hormone replacement. In patients with combined adrenal and thyroid dysfunction due to immune checkpoint inhibitor-related hypophysitis, thyroid hormone replacement should begin 3–5 days after initiation of glucocorticoid replacement in order to avoid an acute adrenal crisis. Endocrine dysfunction following immune checkpoint inhibitor-related hypophysitis is generally irreversible, and high-dose glucocorticoids should only be considered in patients with significant mass effects or severe hyponatraemia. In terms of primary thyroid dysfunction, subclinical hypothyroidism (with normal free thyroxine [FT4] and thyroid stimulating hormone [TSH] <10 mIU/L) could be managed with careful observation and close follow-up. Thyrotoxicosis is typically transient (ultimately giving way to primary hypothyroidism), and may be treated with beta blockers for a brief duration. A radioiodine scan may help to differentiate thyrotoxicosis related to thyroiditis from hyperthyroidism related to Graves’ disease. However, one should be cognisant of the fact that such patients frequently undergo iodinated contrast-based imaging for their primary disease. This may interfere with the uptake of radioiodine by saturating the thyroid gland iodine pool.

**Tyrosine kinase inhibitors**

Tyrosine kinase inhibitors (TKIs) are one of the most potent classes of targeted therapy drugs, used against many different cancer types. They inhibit the enzyme tyrosine kinase, which is involved in the transfer of phosphate from adenosine triphosphate to tyrosine residues in the catalytic domain of growth factor receptors. By this mechanism, TKIs inhibit vascular endothelial growth factor receptors (VEGFRs) and their downstream targets, and suppress endothelial proliferation. The prevention of vascular growth in the tumour causes disruption to the supply of nutrients and oxygen and kills the tumour cells.

**Indications and thyroid dysfunction associated with tyrosine kinase inhibitors**

Currently, the FDA has approved more than 20 different TKIs for clinical use. In comparison to traditional cytotoxic antineoplastic agents, TKIs have high selectivity, high efficacy, a low number of side effects and have superiority in the treatment of chronic myeloid leukaemia, NSCLC and RCC. They are also active against medullary thyroid cancer and differentiated thyroid cancer refractory to iodine-131 (I-131), gastrointestinal stromal tumour (GIST), pancreatic neuroendocrine tumours and hepatocellular carcinoma. Thyroid dysfunction is recognised as an important but manageable side effect of TKIs. Although all TKIs act through the same mechanism, they differ in their spectrum of targeted kinases, thereby resulting in varying rates of thyroid dysfunction. TKIs can induce de novo hypothyroidism which can be preceded by a phase of transient thyrotoxicosis in 20–40% cases. The TKIs which are known to induce new-onset hypothyroidism in a significant proportion of patients include sunitinib, sorafenib, axitinib, pazopanib, vandetanib and motesanib. The other variety of thyroid dysfunction induced by TKI is increased levothyroxine (LT4) requirements in patients who have been on stable doses of thyroid hormone replacement following thyroidectomy for other reasons.
Sunitinib-induced thyroid dysfunction

The TKI most frequently associated with development of new onset hypothyroidism is sunitinib.34,35 It is approved for the treatment of RCC and GISTs resistant to therapy with imatinib. Hypothyroidism occurs in 14–70% of recipients in different studies.21–3 A meta-analysis of seven randomised trials which included 2,787 subjects revealed a risk ratio for all and high-grade hypothyroidism of 13.95 and 4.78, respectively.24 The risk of developing hypothyroidism increases with time and with the number of cycles of therapy.32 The time for development of thyroid dysfunction in the two largest series with long follow-up was found to vary from as early as 4 weeks to as late as 92 weeks.25,26

Thyroid dysfunction induced by other tyrosine kinase inhibitors

Sorafenib is used for the treatment of patients with metastatic RCC, advanced hepatocellular carcinoma and radioactive iodine-resistant advanced thyroid cancer. The incidence of thyroid dysfunction with sorafenib is much less when compared to sunitinib, ranging between 6.3–27.0%.20,24 The median time to develop hypothyroidism was 20 months, but an increase in TSH could appear as early as 6 weeks after initiation of treatment.20,26 In the phase III AXIS trial, axitinib, used for treatment of RCC, caused hypothyroidism more commonly compared to sorafenib (21% versus 7%).20 Pazopanib, a TKI approved for the treatment of RCC and soft tissue sarcoma, has reported rates of hypothyroidism of around 12% or less.31 Other TKIs linked with hypothyroidism include cabozantinib, nilotinib, dasatinib, erlotinib, gefitinib, lapatinib, nintedanib, regorafenib and tivozanib.27

Mechanism of tyrosine kinase inhibitor-induced hypothyroidism

The pathophysiology behind the development of hypothyroidism is presumed to be vascular, resulting from its anti-angiogenic effect. The thyroid gland is a highly vascular organ and its blood flow is mainly dependent on VEGFR signalling. Broad spectrum TKIs like sunitinib, which not only inhibits VEGFR2 and VEGFR3, but also VEGFR1 and the platelet-derived growth factor receptor (PDGFR), impairs vascularisation of the thyroid and induces thyroid ischaemia.3,32 However, PDGFR signalling does not play a role in thyroid angiogenesis in physiological conditions, but becomes active in ischaemia. The compensatory mechanism to restore vascularity might get impaired when it is inhibited.31 The differential effect of various TKIs in inducing thyroid dysfunction can be attributed to their selectivity to block diverse vascular growth factor signalling pathways. A rapid reduction in thyroid vascular flow can result in ischaemic thyroiditis and produce the preceding transient thyrotoxic phase.39 Interestingly, hypothyroidism resulting from TKIs has been associated with longer survival for unclear reasons.36

Tyrosine kinase inhibitors in thyroidectomised patients

Another type of thyroid dysfunction occurs in thyroidectomised patients who have a worsening of stable hypothyroidism and increased LT4 requirement after initiating TKI therapy. Imatinib, currently used in the treatment of chronic myeloid leukaemia, GISTs and other haematological cancers, is most commonly associated with this phenomenon. Imatinib does not induce hypothyroidism in those with an intact thyroid gland.20 Sorafenib and sunitinib have also been reported to cause similar elevation in TSH in patients with pre-existing hypothyroidism.34,35 The possible mechanisms could be stimulation of type 3 deiodinase activity leading to increased peripheral inactivation of thyroid hormones and a dose-dependent inhibition of thyroid hormone transport protein, monocarboxylate transporter.36,38 Additionally, reduced pituitary type 2 deiodinase activity can cause intracellular depletion of triiodothyronine (T3) in the thyrotrhops and an inappropriate elevation of TSH for the concomitant serum free T3 and FT4 levels.39

Screening and management of tyrosine kinase inhibitor-induced thyroid dysfunction

Dru et al. recommend screening by measurement of TSH at the initiation of TKI treatment and then monthly (or on the first day of a new cycle in cases of interrupted treatment) for the first 6 months. Thereafter, TSH estimation should be done every 2–3 months or in the event of clinical signs of thyroid disorder. In individuals with pre-existing hypothyroidism, TSH should be monitored monthly for the first 3 months, followed by monitoring at 3-monthly intervals throughout the therapy period. LT4 should be initiated if TSH is >10 mIU/L or if TSH is between 5–10 mIU/L on two assays, along with clinical symptoms, presence of anti-thyroid antibodies or ultrasound evidence of autoimmune thyroiditis. After the end of TKI treatment, a trial of withdrawal of LT4 should be considered with appropriate monitoring.38

Other immunomodulators

Interferon-α

Interferon-α (IFN-α) is a cytokine approved for treatment of hepatitis C virus (HCV) and several kinds of malignancies like melanoma, RCC, Kaposi’s sarcoma. It is also used for the treatment of hairy cell leukaemia and follicular lymphoma. In recent years, it has largely been replaced as an anticancer agent by better alternatives. It exerts a direct antitumour effect and also indirectly causes the immune-mediated destruction of tumour cells by inducing the expression of major histocompatibility complex-1 (MHC-1), tumour-specific antigen and adhesion molecules on the cell surface.40

The mechanism of IFN-α-induced thyroid dysfunction (IITD) is not fully understood. The expression of MHC-1 antigens on the cell surface results in the activation of cytotoxic T cells, which in turn causes cellular destruction.41 The presence of pre-existing intrathyroidal lymphocytes thereby increases the susceptibility for the development of IITD. This also explains the finding of a higher likelihood of the development of thyroid dysfunction in individuals with anti-thyroid antibodies.42 Mandel et al. classified IITD in patients with HCV into two groups: autoimmune and non-autoimmune.43 Oborzycky et al. suggested the addition of a new term, ‘undifferentiated IITD’, in view of the variable course of the disease.44 The full spectrum of IITD described in patients with HCV may be seen in those receiving IFN-α therapy for malignant conditions.45

Clinical thyroid disease is found in 15–20% of patients with HCV receiving IFN-α, and up to 40% of those become thyroid-antibody positive.46,47 Autoimmune hypothyroidism is the most common clinical presentation, occurring in 20% of cases, followed by destructive thyroiditis (2–3%). Graves’ disease is found rarely.48 Notably, HCV infection itself can induce thyroid dysfunction. IFN-α treatment for malignancy carries a lesser chance of the development of thyroid dysfunction compared with patients with HCV.49 Thyroid disorders occur in 2.4–31.0% of patients receiving IFN-α therapy for solid tumours.5 It is recommended to screen for thyroid dysfunction with TSH and antithyroid antibodies prior to starting IFN-α. Follow-up TSH should be done every 2 months in those with positive antithyroid antibodies and every 6 months in those with negative antibody status.1

Interleukin-2 (aldesleukin)

Interleukin-2 (IL-2) is another cytokine-based therapy used for the treatment of metastatic melanoma and RCC. Its use has declined in recent
years due to the increased availability of better-tolerated alternatives. IL-2 destroys tumour cells by activating NK cells and antigen-specific T cells.43

In a large cohort of 281 patients treated with IL-2 alone, hypothyroidism was reported in 35% of patients, although only 9% of patients required LT4. Longer duration of therapy correlated with a higher risk of development of hypothyroidism. Thyrotoxicosis occurred in 7% of patients receiving high-dose IL-2. The overall incidence of thyroid dysfunction did not differ in the high- and low-dose IL-2 arms. In data from PROCLAIM44, a registry of patients receiving high-dose IL-2, 70% of patients developed primarily vitiligo (in cases of melanoma only) and/or thyroid dysfunction (with a greater incidence in patients with RCC compared with patients with melanoma). The development of these immune-related adverse events correlated with improved tumour control and overall survival.53 Hypothyroidism usually ensues by 4–17 weeks after starting IL-2 and may be reversible on discontinuation of treatment.52,53 The frequency of thyroid dysfunction is 10–60% in various reports, but in many of these studies patients received IL-2 in combination with IFN-α, lymphokine-activated killer cells, or vaccine.44

IL-2 stimulates autoreactive lymphocytes and induces thyroid autoimmunity. It also increases IL-1, tumour necrosis factor-alpha (TNF-α), and IFN-γ. These cytokines induce the presentation of human leukocyte antigen class-II (HLA-II) and associated autoantigens on thyrocytes, and causes its autoimmune destruction.45–48 High levels of thyroid antibodies prior to treatment amplified the risk of IL-2-induced hypothyroidism.53 TSH should be measured before initiating treatment with IL-2 and then once every 2–3 months during therapy.1

Alemtuzumab

Alemtuzumab is a recombinant humanised monoclonal antibody that is directed against the cell surface glycoprotein, CD52, present on lymphocytes. It causes profound lymphopaenia by causing complement-mediated lysis of these cells. The change in immune repertoire that occurs during subsequent lymphocyte reconstitution accounts for its therapeutic effect.46 It is administered for the treatment of T-cell prolymphocytic leukaemia and chronic lymphocytic leukaemia (CLL). Alemtuzumab is also approved for the treatment of active relapsing-remitting multiple sclerosis (MS). It is used as an immunosuppressive agent following solid organ and stem cell transplant and in graft versus host disease.

Thyroid dysfunction commonly occurs after therapy with alemtuzumab for MS, but has not been reported after treatment of malignant conditions.57–60 In a recently published study, thyroid dysfunction occurred in 41% of 248 patients with MS treated with alemtuzumab. Graves’ disease accounted for 72% of those patients who developed thyroid dysfunction, with a median onset of 17 months following the last alemtuzumab dose, and the majority (89%) within 3 years.58 In another large series of 334 patients with MS, thyroid dysfunction was reported in 34% of patients receiving alemtuzumab compared with 6.5% receiving IFN-β1a. Graves’ disease was reported in 22%, hypothyroidism in 7%, and subacute thyroiditis in 4%. Thyroid-binding inhibitory immunoglobulin was positive in 74% of patients with overt hypothyroidism.59 Patients receiving alemtuzumab for T-cell prolymphocytic leukaemia and chronic lymphocytic leukaemia did not develop thyroid dysfunction.60–63 Autoimmunity following alemtuzumab therapy has been attributed to a breakdown in self-tolerance during immune reconstitution and is mediated by humoral mechanisms arising from homeostatic T-cell proliferation.64 The absence of thyroid dysfunction in neoplastic conditions in comparison to MS is presumed to be due to the simultaneous use of other immunosuppressive agents in patients with cancer, or to the tendency for underlying autoimmunity in patients with MS.1

Thalidomide analogues

Thalidomide and its analogues lenalidomide and pomalidomide are used for the treatment of multiple myeloma, myelodysplastic syndrome and mantle cell lymphoma. The antineoplastic effects are mediated by antiangiogenic, antiproliferative and immunomodulatory activity.42 The incidence of subclinical hypothyroidism is 20% and overt hypothyroidism is 7% during the first 6 months of thalidomide therapy.41 In a retrospective series of 170 patients receiving lenalidomide, hypothyroidism was reported in six, and thyrotoxicosis in four.44 In another series, lenalidomide caused hypothyroidism in 25.8% of cases after a median duration of 5.2 months in patients with diffuse large B-cell lymphoma.46 Two case reports of pomalidomide-induced hypothyroidism have been described in the literature.44,45 Slean and Silkiss reported a case of lenalidomide-induced eyelid retraction in a 76-year-old woman who was euthyroid but had elevated thyroid antibodies.46

One of the probable mechanisms behind the development of hypothyroidism is the induction of ischaemic thyroiditis through the antiangiogenic effect of thalidomide. The deregulation of cytokines or direct effects on T-lymphocytes have also been hypothesised to trigger an autoimmune response to the thyroid.47 Elevated anti-thyroid peroxidase antibodies and increased TNF-α levels have been demonstrated in these patients, further corroborating the possibility of immune-mediated pathogenesis.60,61 Additionally, lenalidomide might cause increased T-cell proliferation and activation by stimulating tyrosine phosphorylation of T-cell costimulatory molecules (e.g., CD28). This may inhibit regulatory T cells and might block immune checkpoints like CTLA-4.46 Measurement of TSH is recommended before initiation of treatment with these agents, and every 2–3 months thereafter for the duration of therapy.22

Radiiodine-based cancer therapy

Metallobenzylguanidine (MIBG) is a norepinephrine analogue. Combined with I-131, it is used as a radiotherapeutic agent to treat tumours derived from neural crest cells. Active transport by uptake-1 system is the dominant mechanism of uptake in these cells, while passive diffusion plays a minor role.52

Indications and adverse effects

I-131 MIBG therapy has been used for a variety of tumours including inoperable pheochromocytoma or paraganglioma, stage III or IV neuroblastoma, inoperable carcinoid tumours and metastatic or recurrent medullary thyroid carcinoma.26 Adequate uptake and retention of MIBG during a pre-therapy scan is a prerequisite before the administration of a therapeutic dose of I-131 MIBG. The adverse effects associated with the therapeutic use of I-131 MIBG include transient hypertension, bone marrow suppression and thyroid dysfunction.23 During its administration, free radiiodide formed due to chemical instability and biological degradation in the liver may result in toxicity to the thyroid gland.

Spectrum of thyroid dysfunction

Thyroid dysfunction following I-131 MIBG therapy has been reported in the form of hypothyroidism, radiation thyroiditis, thyroid nodule and thyroid carcinoma.53–56 Due to higher radio-sensitivity, the concern remains greater in children receiving this therapy, most commonly for neuroblastoma. Traditionally, when using iodine-123 or I-131 MIBG for diagnostic or therapeutic purposes, thyroid protection is rendered using a saturated solution of potassium iodide or Lugol’s iodine.57 Despite the use of thyroid protection, the incidence of thyroid dysfunction following MIBG therapy is quite high, and varies from 52–86% across studies.23–28
Table 2: Recommendations for the screening and management of thyroid disorders during anticancer therapy

<table>
<thead>
<tr>
<th>Drug</th>
<th>Screening</th>
<th>Management</th>
</tr>
</thead>
<tbody>
<tr>
<td>Immune checkpoint inhibitors</td>
<td>TSH and FT4 before initiation, should be repeated before each cycle</td>
<td>Primary hypothyroidism: start standard LT4 therapy: initial full dose (1.6 mcg/kg) in young and healthy; reduced dose in elderly or patients with cardiovascular disease</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Thyroiditis: usually self-limited, beta blockers may be given for a short duration</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Graves’ disease: standard therapy</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Central hypothyroidism due to hypophysitis: start LT4 at 1 mcg/kg, adjust dose by monitoring FT4 every 6–8 weeks, rule out or treat associated hypocalciositism before initiation</td>
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<tr>
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<td>I-131 MIBG therapy</td>
<td>TSH and FT4 at baseline and at every 3–6 months</td>
<td>Primary hypothyroidism: LT4 therapy if TSH level &gt;10 mIU/L. Some authorities recommend treatment even with TSH 5–10 mIU/L, because of theoretical risk of malignancy due to radiation</td>
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<td>Prior hypothyroidism: increased levothyroxine requirement as judged by TSH value</td>
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<tr>
<td>Bexarotene</td>
<td>TSH and FT4 at initiation of therapy</td>
<td>Central hypothyroidism: start LT4 at 25–50 µg along with bexarotene. FT4 levels every 1–2 weeks for first 4 weeks; dose adjusted to maintain it at upper third of normal. Monthly FT4 once levels stable. Treatment discontinued on stopping bexarotene.</td>
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<td>Prior hypothyroidism: increased LT4 requirement, and can increase 2–3 times; should be adjusted according to FT4 levels</td>
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</tbody>
</table>

**FT4** = free thyroxine; **I-131 MIBG** = iodine-131 metaiodobenzylguanidine; **LT4** = levothyroxine; **TPO** = thyroid peroxidase; **TSH** = thyroid-stimulating hormone.

### Incidence of thyroid dysfunction

In a study by van Santen et al., TSH elevation (>4.5 mIU/L) was reported in 22/42 (52%) subjects with neuroblastoma with a mean follow-up duration of 1.4 years following MIBG therapy. In another study by the same group, long-term thyroid outcomes among 25 survivors of neuroblastoma were reported. After a median follow-up of 6 years, 14/25 (56%) subjects had persistent TSH elevation, while 6/25 (24%) developed thyroid nodules or cysts. In another study by Picco et al., a follow-up of 6–12 months following MIBG therapy, TSH elevation was evident in 12/14 (86%) survivors of neuroblastoma. Of these, eight (67%) had overt hypothyroidism, while four (33%) had compensated hypothyroidism. In a study comparing two thyroid protection strategies, a combination of potassium iodide, thyroxine and methimazole (cocktail protection) was compared with potassium iodide (standard protection) in children with neuroblastoma receiving therapeutic I-131 MIBG. The study evaluated 23 children who were given the cocktail protection (beginning 1 day before diagnostic I-131 MIBG and continued until 4 weeks after therapeutic I-131 MIBG), and were followed up for a mean duration of 19 months. The control group was historical cohort of children with neuroblastoma (N=42) who received standard thyroid protection. Thyroid function was normal in 86% survivors in group A (cocktail protection) compared to 44% in group B (standard protection). The authors thus concluded that a combined protective strategy may be more effective against radiation damage in patients receiving MIBG therapy for neuroblastoma. The long-term efficacy of this strategy, however, remains to be seen.

### Screening and management

Although a clear recommendation is lacking, it is advisable to obtain a thyroid function at baseline and at 3–6 monthly intervals in patients receiving therapeutic I-131 MIBG. LT4 supplementation is recommended in those with low T4 levels or TSH level >10 mIU/L. However, considering the theoretical concern of increased neoplasia risk with chronically elevated TSH levels, some authors recommend treating even mild subclinical hypothyroidism (TSH <10 mIU/L) in the setting of previous radiation exposure. Since thyroid radiation is a known risk factor for malignancy, and increased incidence of thyroid nodule has been reported in studies, such patients should be followed indefinitely for the same.

**Bexarotene**

Topical and systemic preparation of bexarotene, a third-generation retinoid analogue, is approved for the treatment of cutaneous T-cell lymphoma. It is a selective agonist of retinoid X receptor (RXR), a member of the nuclear receptor superfamily. RXR functions by forming heterodimer with thyroid hormone receptor and other nuclear receptors. T3 exerts a negative feedback effect on the transcription of the β-subunit of TSH after binding to its receptor and subsequent heterodimerisation with RXR.

**Mechanism of thyroid dysfunction**

Bexarotene alters the feedback effect of thyroid hormone on the pituitary by causing reversible thyroid hormone-independent inhibition of TSH gene expression. It blocks transcription even in the absence of T3, and decreases TSH production. Besides, bexarotene causes direct inhibition of TSH secretion, and a rapid fall in serum TSH occurs even after the administration of a single dose.

**Central hypothyroidism**

Oral bexarotene administration has been found to induce rapid onset and reversible central hypothyroidism. In trials with more than 20 patients on bexarotene, the percentage of patients developing central hypothyroidism varied from 29–100%. In a recently published study from Japan, hypothyroidism developed at 1 week in 45/66 patients, and only five patients remained euthyroid at 1 month. Serum TSH levels returned to normal within 1 week of discontinuation of bexarotene in 90% of cases who were euthyroid earlier.
The factors predicting the development of central hypothyroidism are higher doses of bexarotene (>300 mg/m²/day) and prior treatment with IFN-α. The aforementioned Japanese study also demonstrated that a pre-treatment FT4 (free thyroxine) value in the lower-normal range or a pre-treatment TSH value <1.30 µIU/L correlated with the development of central hypothyroidism. Iodine deficiency and polymorphism of the RXRγ gene were also proposed as other possible determinants of the development of central hypothyroidism. RXRγ and the negative thyroid hormone-responsive element are both present in the promoter region of the gene encoding the TSHβ-subunit, and bind to retinoid and thyroid receptors respectively to inhibit its transcription.

Other effects on thyroid metabolism

Bexarotene also exerts peripheral effects by increasing the degradation of thyroid hormones. Ten post-thyroidectomy patients with pulmonary metastases of differentiated thyroid carcinoma who received a 6-week reinduction course of bexarotene had a drastic fall in serum total T4, FT4 and total T3 levels. The increased catabolism of thyroid hormone was thought to be independent of peripheral deiodinase activity and likely to be mediated by other pathways like glucuronidation and sulfation.41

Management

The recommendations suggest initiation of LT4 at 25–50 µg per day when bexarotene treatment is started.95–7 FT4 levels should be monitored every 1–2 weeks for first 4 weeks and adjusted to maintain FT4 within the upper third of the local normal laboratory range. There is no role of monitoring serum TSH levels. Once two consecutive blood reports are available, frequency of monitoring can be decreased to once every 1–2 months. In many cases, the thyroid dysfunction is temporary and treatment can be withdrawn with discontinuation of anticancer therapy.

Conclusion

Immune-based and targeted chemotherapeutic agents can produce a distinct group of immune-related adverse events, foremost among them being thyroid dysfunction. Appropriate screening for primary thyroid dysfunction should be performed when immune checkpoint inhibitors, TKI, IFN-α, IL-2, thalidomide and other immunomodulatory drugs are administered. Guidelines and recommendations on the management of thyroid disorders induced by common anticancer drugs are summarised in Table 2.37,10,7,27–31 Central hypothyroidism is also described as a common adverse event with bexarotene and immune checkpoint inhibitors, albeit through different mechanisms. Administration of LT4 should be instituted when appropriate. In many cases, the thyroid dysfunction is temporary and treatment can be withdrawn with discontinuation of anticancer therapy.
Anticancer Drug-induced Thyroid Dysfunction


